

Save our Flora

AN ONLINE INDEPENDENT NATIONAL PROJECT Conservation through Cultivation

Project launched on

14th November 2013

Maria Hitchcock Administrator
Bulletin Editor

Bob Ross Conservation
Legislation

Membership Individuals: 145
Groups: 19
International 2

Membership is free.

Please encourage others to join.

Bulletins are sent by email only.

Feel free to pass them on.

People joining up after e-Bulletin No 5 is published will receive the latest e-Bulletin only. Earlier Bulletins can be sent out on request. This is an informal interactive sharing group. We welcome your emails, articles and offers of seed and cuttings at any time.

Your privacy is respected and assured with this group.



Caladenia lindleyana Tas Critically Endangered

Image: www.retiredaussies.com

**You can now access all our
previous E-Bulletins online**

Go to

**[http://coolnatives.com.au/
SaveOurFlora.html](http://coolnatives.com.au/SaveOurFlora.html)**

In this issue:

From the members	2-3
National Threatened Species Day	4
Grafting Native Plants	5-12
Ministers Letter	13-14
Seed & Cuttings Exchange	15
Protocols for requesting seed and cuttings	16

Unsure if you have any rare or endangered plants? Check them out on

Save our Flora

From the members:

Ruth Crosson (Gladstone) writes:

Correction about the *Attalya*. Brent informed me that what I have, rare and threatened is *Atalya collina* Yarwun Whitewood. *A. calciola* is now off the list of rare and threatened plants.

A. collina is what is on Boyles Property, Boyles Road Yarwun. Seed was collected by Ted Johansen, when he rented the farm house. He germinated some seeds and gave me 3 plants. This was before it was identified by Qld herbarium and named *A. collina*. Only one survived and is now about 3 m tall, planted on council street, near cnr Short and Goondoon, near a water pipe line, which runs down Short St.

Note: This species is known from only two populations in Queensland: west of Gladstone at Yarwun and near Ubobo, west of Miriam Vale (Queensland Herbarium 2008a). The Yarwun population consists of 10 trees of an even age structure, with all plants 3-4 metres tall and no sign of regeneration at the site (Barry & Thomas 1994).

http://www.environment.gov.au/cgi-bin/sprat/public/publicspecies.pl?taxon_id=55417

Ed: Do you know if there is any viable seed on the surviving plant, Ruth?

This Conservation Advice was approved by the Minister / Delegate of the Minister on: 16/12/2008

The main identified threats to *A. collina* are its low numbers and lack of long term security. At the Yarwun site the 10 trees occur in a partially cleared and highly disturbed site subject to cattle grazing (Barry & Thomas, 1994). The southern population also occurs on partially cleared land. The main potential threats to the species include the effect of grazing, weeds and fire protection (Barry & Thomas, 1994). The ecological requirements and reproductive needs of *A. collina* are not known.

Enable Recovery of Additional Sites and/or Populations

- Undertake appropriate seed collection and storage.
 - Investigate options for linking, enhancing and establishing additional populations.
 - Implement national translocation protocols (Vallee et al., 2004) if establishing additional populations is considered necessary and feasible.
- <http://www.environment.gov.au/biodiversity/threatened/species/pubs/55417-conservation-advice.pdf>



Attalaya collina - Endangered EPBC list
Herbarium specimen collected on M. Boyle's property in 1990 and in the Kew Gardens collection.
For a larger view and more detail go to

Save our Flora

From the members:

Kerry Rathie (Toowoomba) writes:

I have been busy finishing off my book on Brachychitons, & settling in to my relatively new garden (3 years old in a month) here in Toowoomba. The book should be printed next month, & I am self-publishing, as my then co-authors & I did with our field guide to the plants of S-E Qld., 'Mangroves to Mountains'. The current book is 220 pages, mainly consisting of colour photos of the 40-odd species & a larger number of cultivars. Most of both will be unfamiliar to almost everyone not in the Brachychiton study group, as most of the cultivars are very newly released or, more often, not yet released, & the majority of species & natural hybrids are from the tropics, In particular from sparsely populated areas of Cape York & the Kimberleys.

While soliciting photos & plant material for the book from my contacts, 3 new natural hybrids emerged from the woodwork, plus several entities of unknown ancestry which may be colour or habit variants of the named taxa, or may not. For most of these I now have 1 to ten grafted plants on hand, but it will be a little while before I can hand out samples. As a geneticist I will be trying to pin down their relationships over the next few years, my health permitting, & hope to collect material from some of the mysteries as well. A bit of DNA work should sort most things out.

Some of the mysteries have been grown in a few Qld gardens for some years, namely the 'all-red' form of *B. garrowayae*, 'Big Pink', 'Coen Pink', 'sp. Mt. White' & 'Argyle Pearl'. The last in the list is a white shrub collected near Lake Argyle, & the 3 preceding it all come from near Coen on Cape York, & are utterly different apart from all being beautiful & strikingly different from southern members of the genus. All the taxa in this paragraph are growing & flowering well in the subtropics & in distinctly cooler Toowoomba, although Argyle Pearl, even when grafted, is very slow-growing in both environments. For all I know, it may also be slow in the E Kimberley. Only one plant was ever seen in the wild. My garden, & almost all of Toowoomba, is virtually frost free, due to the Urban Heat Island effect & to cold air sliding down the nearby slopes, but is prone to very windy weather quite often, & many cold days in autumn & winter just a half degree or so above freezing. Some orchids & grevilleas get cold burns despite no actual frosts.

I have quite a few fairly unique plants here, but in most cases I only brought one or two specimens of each, & have not had time until now to do any multiplication. I have Arnhem Land Wilkea spp., for instance, several Citrus species (were Microcitrus), & a range of fern mutant forms like silver Pellaea paradoxa & numerous crested & forked etc Drynarias & Doodias, & some unusual forms of Pandorea or Tecomanthe. The little red Pandorea sp. Mt Maroon has become a Tecomanthe, but I am not sure how many of the others have been transferred. Have just struck a batch of what was P. sp. Theodore, a nice purple inside & cream outside form.

Margaret Lee (Adelaide) writes:

Thank you all so much for your interest and help regarding finding a *Eucalyptus deglupta* tree to photograph in Australia. One of the wonderful things about ANPSA is the willingness to help and share our passion for plants.

There is possibly a mature one at Edmonton, near Cairns. I've yet to find someone to check it out. According to one of the websites there are very young ones in Melbourne Botanic Garden, but they'd be too young to photograph.

Ed: Had a look at some Google images of E. deglupta. It is astonishing - the most gorgeous colours. Here's a photo.



Image: www.organichealthalliance.com

Save our Flora

National Threatened Species Day

National Threatened Species Day, held each year on 7 September, aims to encourage the community to prevent further extinctions of Australia's fauna and flora, and to restore healthy numbers of threatened species and ecological communities in the wild. This is a time when many Australians celebrate our unique and valuable biodiversity with activities to protect and conserve the environment.

National Threatened Species Day was first held in 1996, to commemorate the death of the last Tasmanian Tiger in captivity in 1936 in Hobart. The concept was developed by the Threatened Species Network, a community based program of the Australian Government's Natural Heritage Trust and the WWF Australia, as a way to showcase Australian threatened species.

By focusing attention on the plight of many of our threatened animals and plants, Threatened Species Day aims to encourage greater community support and hands-on involvement in the prevention of further losses of Australia's unique natural heritage.

A number of events are coordinated across Australia for National Threatened Species Day to raise community awareness about the plight of threatened species in Australia and to encourage community participation in conservation activities. Activities range from exhibitions and festivals, to displays, guided walks, workshops and a range of other community education and promotional activities.

There is an educational activity sheet suitable for teachers and students available on <http://www.environment.gov.au/system/files/resources/3367c4a6-4e79-4677-b6fb-d382f5808598/files/tsdo6time-act.pdf>

Forward it on to a teacher friend. However, some of the ideas can be implemented by individuals and could be included in newsletters and bulletins. Become a threatened species ambassador.

Something I've been doing is creating habitats in my garden. Here's the item from the pdf.

Critically endangered

Caladenia actensis ACT
 Caladenia anthracina Tas
 Caladenia campbellii Tas
 Caladenia cremna Vic
 Caladenia intuta SA
 Caladenia lindleyana Tas
 Caladenia lodgeana WA
 Caladenia melanema WA
 Caladenia pallida Tas
 Caladenia procera WA
 Caladenia pumila Vic
 Caladenia saggicola Tas
 Caladenia sp. Kilsyth South Vic
 Caladenia sylvicola Tas
 Caladenia tonellii Tas

Hibbertia priceana WA
 Hibbertia puberula subsp. glabrescens NSW
 Hibbertia tenuis SA

Isopogon robustus WA

Lomatia tasmanica Tas

'Create habitat for native animals and birds: Plant local native species in your garden and add logs and rocks to create habitat for native animals and birds. It is important to plant native species local to your area, as some plants can become serious weeds and end up doing more harm than good'.

Ed: I've now planted out a large number of Lomandra longifolia which will become a lizard lounge. In another part of the garden I planted a big clump of paper daisies to make a butterfly bar. I'm sure you can think of all sorts of catchy names for micro-environments like this. Why not share them with us.

Ref. <http://www.environment.gov.au/resource/protecting-australias-threatened-species>



Save our Flora

Grafting Native Plants

by Merv. Hodge¹
Brisbane, Qld.

¹As a preface to this article I would like to pay tribute to the late Harvey Shaw who first interested me in grafting Australian native plants. He gave assistance and encouragement to a number of people interested in developing this skill.

Why do we need to graft plants? The main reason is to provide a reliable and vigorous root system not susceptible to root diseases and able to perform satisfactorily in local conditions. In addition, the scion material will produce a plant with identical flowering and fruiting habits to the parent. In rare cases it could be used to prevent undesirable suckering habits of the parent plant. It may also be used where a plant is too difficult to propagate by other means. It is a more labour-intensive and costly means of producing plants, so the end product is necessarily more expensive. This cost may vary from plant to plant, depending on the degree of difficulty.

Grafting of Australian native plants is not new, some having been successful at least early this century – e.g. grafting *Correa* on *C. alba* and *Clianthus formosus* onto *C. puniceus* (a New Zealand species).

Selection of Plant Material

Rootstocks need to be closely related to the scion, i.e. they need to be in the same family and preferably in the same genus. This is no guarantee of compatibility and ultimately it comes to actually trying them and observing the results over a long period. It is probable that different combinations of scion and rootstocks will be necessary in different climatic regions.

A critical assessment of the rootstock available will soon eliminate all but a few, perhaps leaving only one. The points to consider are compatibility, reliability, longevity and availability. It might be better to select a reliable, easily propagated hybrid rootstock over an equally compatible species which may be difficult to propagate or available only seasonally. Rootstocks should be healthy and vigorous, both before and after grafting. Any stress placed on the rootstock can cause it to abort the graft even though the rootstock might survive and look quite healthy. This can be caused by something as simple as overlooking the watering of a plant on one occasion. Good potting mixes and fertilizers are important to maintain vigour. A significant factor in my success rate is vigorous tubed plants of *Grevillea robusta* from Fairhill Nursery.

Scion material is best taken from a clean disease-free, vigorous plant, in the same manner as when selecting cutting material. Better results are achieved using scions taken from healthy potted plants, rather than those collected from garden plants or from wild sources. Harvey Shaw preferred soft green tips for scion and rootstock for his grafts. I prefer firm but green material. Both work equally well, but I find the latter easier to work for my method.

Grafting Methods

There are a number of different methods of grafting. The methods most commonly used are top wedge, whip and approach grafts. The latter was that which Harvey Shaw mostly used until he perfected his technique with top wedge grafts. I have only used approach grafts on a few occasions and will not describe it here. I used top wedge grafts for some time, but later switched to whip grafts.

Vol. 15—Page 369

Save our Flora

The Top Wedge Graft

For the top wedge method, select scion material and cut to lengths of 25 to 75 mm long. Compare the scion to the rootstock to determine a section of equal diameter where the graft will be made on the rootstock. Remove the top of the rootstock, cutting directly below a node on green wood. I prefer to do this to remove any possibility of the rootstock producing laterals at the node. I find no disadvantages in doing this, despite predictions of die-back that others have quoted.

The scalpel is then placed along the diameter of the stem (the other hand may have to hold the rootstock steady). The blade is then pushed firmly and carefully vertically to form a slit down the stem, effectively dividing it into two equal portions, to a depth of 18 to 25 mm. Care should be taken not to allow the blade to wander off course. To do so could cause a disaster to the graft and to the hand holding the rootstock. I must admit that it is this part of the operation which first caused me to seriously consider using the whip graft.



Now a wedge is cut on the end of the scion, approximately the same length as the slit made in the rootstock. The wedge should be made by making an angled cut on each side of the scion to meet at the original diameter. Firmly, but carefully, insert the wedge into the slit made in the rootstock until the tip of the wedge has penetrated to the full depth of the cut. Care should be taken that the two stems line up so that the cambium layers are making as much contact as possible. In most cases there is enough force exerted by the graft to hold the scion in place whilst hands are moved to attach the tape. The tape should be bound firmly over the graft in two layers, firmly stretching the tape so that it holds the graft together and seals it against the outside air and moisture.

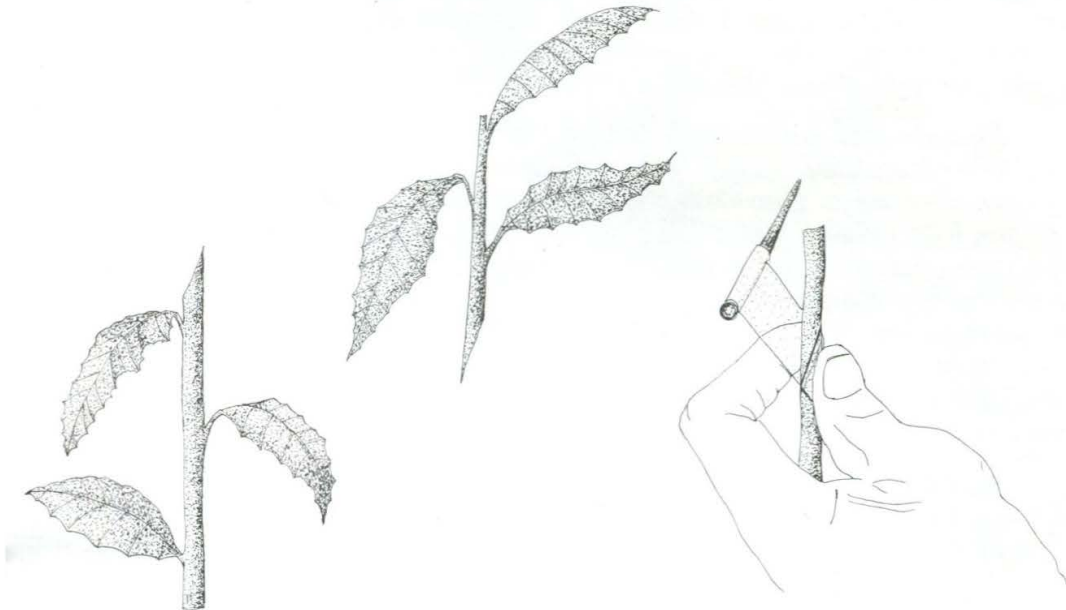
Notes on the Top Wedge Graft — All plants are a little different. The thickness of material indicated would be as for *Oreocallis*. It is thicker than I would generally use for say *Grevillea*. The length of most scions (upper plant) that I have done recently have been 5-13 cms long, again a little longer than those indicated. The most important condition was to ensure that they had at least a few good, potential growing points, i.e., at least three terminal and/or lateral buds. Suggestions as to the tools and tape to use are at the end of the article.

Save our Flora

The Whip Graft

The whip graft is the method which I now prefer and I now do all grafts that way. I find it simpler and faster and just as effective as wedge grafts. Whilst it is claimed that wedge grafts are stronger, I have not found any significant difference between the two once the grafts have taken. Both have their share of mechanical failures and both exhibit good strength if the graft is well done. I find that I can do whip grafts faster, but it requires more dexterity to hold the graft together whilst starting the tape. I do not find it necessary to add a tongue as in the “whip and tongue” method. That would slow up the process and make matching of cambiums more difficult.

Firstly, ensure that you have tape wound on two nails (see page 374). The end of the tape should be left with about 10 mm unwound. Then begin the whip graft by cutting the top off the rootstock below a node. This type of graft requires an acute angled cut across the entire thickness of the scion and rootstock, starting at a point on each where their stem diameters are about equal. The cut should be made at the same angle for both and continued until it completely severs the unwanted portions of the scion and the rootstock. Ideally, it should be completed with one cut 20 to 30 mm long, and should end in a fine tip. If this is done correctly, the cambium layers should appear identical.



The scion and rootstock should be placed together so that their cambiums match and held firmly with the left hand towards the bottom of the graft. The right hand should then pick up the nail with the tape already wound on it. Place the unwound section of the tape in a position where a slight movement of the left thumb will allow it to press the tape against the rootstock side of the graft.

Notes on the Whip Graft to this point: Perhaps it may help to comment on a few aspects to this point. The beautiful sketches above provide the key to success. The scion or upper plant may be longer, possibly with more potential growing points. As shown over page, the top cutting may even have the terminal leaves. The material used is often thinner making the job a little more difficult. The grip on the whip graft is not quite right (even though it was made from a photograph of myself in action). The index finger should be a little lower to allow the tape to be initially wrapped around the top of the graft, but high enough to hold the graft together. The tape is initially clamped against the stem by the thumb as shown.



Using a nail, now wind the tape onto the top part of the graft, pulling firmly to hold the graft in place. Gradually slide the left hand down to hold the bottom of the graft. Quickly check to see that the cambiums are still aligned and continue to wind the tape downwards around the graft. Continue the process until the tape fully covers the graft then wind another layer of tape back up the graft, ensuring that the tape does not twist. The tape may be pulled tight at the end of the graft where it will seal itself and break off. This is the most difficult part of the method and it does require some practice. If you are unable to master this, it may be necessary to do wedge grafts. Left-handed persons would have to adapt the description to their own needs.

Once the graft is completed it is necessary to prevent the scion from dehydrating. This can be achieved in a number of ways. You may devise your own method, but the following methods have been well tried and do work.

Dehydration prevention, Method 1: I followed the method used by Harvey Shaw. Some small lateral growths and foliage can be left on the scion, but it is wise not to overdo this. Purchase self-sealing bags about 120 mm x 74 mm with a built-in zipper-like device. Place the bag over the scion and seal it on or below the graft. I make a slight kink in the zipper where it contacts the stem to reduce its potential to open itself. Once the graft has taken the bag must be gradually opened over a number of days to allow the scion to harden off. Grafts can be lost at this stage. I still use this method when the scion has dense, fine foliage which would take too long to remove, or where the scion has no dormant buds and has already produced lateral growths.

Dehydration prevention, Method 2: If the scion has dormant buds, particularly when they appear to be about to grow, and if the plant has easily removable foliage, then I use the following method. Remove all foliage except for small tip leaves. Make the graft as previously described, but do not break the tape once the graft is fully bound. Now continue the tape right up, binding over the entire scion except for the very tip. If the growing tip has previously been removed, then the entire scion can be bound. This should be done with one layer of tape only, stretching it as you bind. Overlap the tape slightly as you bind to ensure that the scion is fully sealed except for the tip. If the tip is very soft it might be best to remove it.

In two to four weeks the buds should become active. They normally break through the tape and continue to grow. As they grow they acclimatize themselves. The tape will eventually decompose and normally does not need to be removed.

I first conceived this idea in 1986 when I noticed that when tape was bound over buds in the scion and rootstock near the graft, they occasionally burst through the tape and continued to grow. I reasoned that if it happened at the graft, it could also occur on the scion. But it would require going against advice from others, i.e. to leave some foliage on the scion. I carried out trials with a number of plants and they were successful.

The photographs opposite illustrate the splice graft and recommended treatment for after care. Photos A, B and C from the bottom up the left hand side show progressive stages of wrapping the scion as for the dehydration method 2. Currently I would not waste time binding the lateral growth on the left but would remove it. The photograph on the top right hand corner shows the two methods of dehydration described here. Currently I would remove the large leaf on the top of the wrapped specimen, allowing only the tip exposed. Leaving the tip does not appear critical if there are good active lateral buds. The two photographs on the bottom right are completed wrapped grafts that have subsequently grown to vigorous plants.



Photographs by Merv Hodge



In March, 1988 I tried this method on a seedling *Grevillea banksii* growing in full sun in the field, using a *G. bipinnatifida* scion. No further assistance was given to the plant. The weather was hot but conditions were favourable to promoting active growth. The scion started to break through the tape in two weeks and continued active growth. On the same day I grafted *Grevillea bipinnatifida* onto two healthy potted *G. robusta* seedlings. These were watered regularly and fertilized and kept under shadecloth. There was a tremendous contrast in the growth rates between the field plant and those in the shadehouse. The field graft was three times the size of the other two in a short time and continued to out-perform them for some time.

After recently having been shown a fogging system in operation, I feel that it would be worth experimenting with grafts in that controlled environment. It would most likely eliminate the need for bags or wrapping. These systems are expensive so the enthusiastic amateur or the small commercial producer may be content to try the methods I have described.

Tools:

Experienced grafters, particularly professionals, have their own preferences. I have tried to improvise with a number of implements, but I eventually followed Harvey Shaw's lead and now use a scalpel with a number 11 blade. It appears to be the optimum for this type of grafting. It is necessary to use a very sharp implement so a good supply of replacement blades is necessary. When the blade starts to pull or tear it is necessary to replace it. The blade should be kept as clean as possible. I pour boiling water over the handle and blade before commencing. I normally dip the blade in Methylated Spirits frequently when grafting. I also use scissors and two nails, see page 374.

Tape:

I prefer to use "Nescofilm" tape, a self-sealing, opalescent, thermo-plastic film. It may be purchased in a 10 cm x 40 metre roll from which I cut strips 10 cm long x 10-13 mm wide. Peel off the paper backing first. This roll will do thousands of grafts. Nescofilm is normally used in laboratories so it should be sought out from medical or scientific suppliers. The scalpel handle and blades can be obtained at the same time. This tape has the advantage of breaking down in sunlight, making it normally unnecessary to remove it after the graft has taken. It remains intact long enough for the graft to take. The tape has a fair amount of elasticity and will give a little with plant growth. It is self sealing and eliminates the need for sealing the finished graft with sealing wax or vaseline.

The tape can be difficult to handle, particularly when grafting small plants. It is inclined to twist and stick together so a method of control is necessary. I found that I could best achieve this by winding the tape onto one end of a match. I later changed to a 65 mm nail after breaking numerous matches. It is advisable to have a second nail on hand with the tape already wound onto it. Inevitably, the tape will break at an inconvenient time when one hand is holding the graft together and the tape has stuck on the nail without a free end. When winding on, the tape should be lightly tensioned and kept flat. If it is allowed to twist and form into a cord-like condition it will tend to strangle the plant, particularly when it is considered that it takes longer to break down in that condition. Too many layers will also increase breakdown time.



Grafting That Has Been Successful

The following are the successful grafts, most of these by Harvey Shaw. All of them are still alive but it may be some time before we can assess their complete success. In most cases there are only small numbers and in some cases only one plant is still surviving. Others are so popular that over a hundred have been grafted and distributed. There are other plants not included which showed initial success and then collapsed months later. These include *Verticordia grandis* onto *Kunzea flavescens* and *Baeckea* 'Winter Pink' onto *Kunzea flavescens*.

Grevillea have been favoured and about 200 forms grafted. Not all have been completely successful, e.g. *G. quercifolia* grafts easily but frequently fails later because it is too brittle at the graft and breaks off. Most have been grafted onto *Grevillea robusta* but a few have been grafted onto *G. banksii*, *G. 'Ivanhoe'*, and *G. 'Royal Mantle'*. Some of the hairy-leaved *Grevillea* responded poorly using the above methods. They appear to be adversely affected by the humid conditions created. I managed to graft a couple without wrapping the scion or using the bag method.

Banksia have proved to be difficult and we have only been successful with a small handful, all onto *B. integrifolia* rootstock.

Hibbertia, including *H. stellaris*, have been grafted onto *H. scandens*. Considering the stem thickness of this plant, grafting it onto anything is astounding. *H. miniata* was successful initially but was lost later.

Eremophila species (14) have been grafted onto the more reliable *Myoporum* species. They grafted well but did not perform well in Brisbane because of the effect of our summer humidity on their foliage.

Prostanthera were grafted onto *Westringia fruticosa*. One notable success was *Prostanthera magnifica*. Most of the *Prostanthera* were found to flower poorly in our conditions so Harvey lost interest in them.

Eucalyptus have been a frustrating challenge. At the time of his death in May, 1989, Harvey was producing promising results grafting good colour forms of *Eucalyptus ptychocarpa* back onto seedlings of the same species. I have been informed that one gentleman has grafted *Eucalyptus ptychocarpa* onto *Angophora costata* in south-east Queensland and that it has been growing well in the ground for some time. He has also successfully grafted *E. ptychocarpa* onto *E. citriodora* and *E. viminalis*, and grafted *E. ficifolia* onto *E. tessilaris*, *E. citriodora* and *E. maculata*. The latter is about 20 years old and still doing well. He has several other combinations of *Eucalypt* scions and rootstocks.

The Geraldton Wax, *Chamelaucium uncinatum* was grafted onto *Kunzea flavescens* and *Bakea virgata* but they were difficult and not always reliable.

Hakea salicifolia has been the recommended rootstock for other *Hakea* species. Whilst many *Hakea* graft readily to that species, it is not reliable enough here in south-east Queensland to be the ideal rootstock. I decided to try grafting *Hakea* onto *Grevillea robusta*. Many have grafted readily to it and, at this stage, it appears to be the best rootstock. The long-term reliability of these is yet to be proved. Another person has grafted a *Grevillea* onto *Hakea salicifolia* because he had no other rootstock on hand. It is still alive.

If you wish to try any of these methods I caution that trials should be carried out to ensure that you perfect the technique for your conditions before committing large numbers of plants to the project.

Save our Flora

Merv Hodge writes:

The article above was published in Australian Plants in 1990 (Vol 15) following my article in the seminar papers of the ASGAP [now ANPSA] of the **14th Biennial Convention Canberra 1988**, under the title of '*A New Method of Grafting Native Plants*'. Actually it applied to exotics as well and is now used commercially by some propagators. The method is acknowledged in '*The Grevillea Book*' by Olde and Marriott Vol 1, Page 135, with further comments on succeeding pages. A supporting photo appears on page 133. You will find many of my photos in all three volumes. I devised this method by observation followed by successful trials. No one else was involved in spite of some speculative writing later.

After later experience with grafting I found that some of the above information needs revision.

I ran a registered Nursery [*Loganview Nursery*] here for about 15 years before and specialised in grafted natives. We closed the Nursery in 2007. During that time I produced thousands [possibly tens of thousands] of grafted natives. It included a range of genera. After this experience I changed my mind on some of the information that I originally published mainly in the combinations of scions and rootstock. I found that there was immediate incompatibility [graft would not take] to long term incompatibility where one way or another the grafted plants were not successful after some weeks or months. I also found that many grafts needed a close relationship between scion and rootstock. All grevilleas are not compatible to one another and similarly all Eucalypts are not compatible to one another. You cannot graft a corymbia to a eucalypt successfully. In eucalypts it is best to select rootstocks from the same subgenus, whereas some myrtaceae can accept intergeneric grafts. E.g. leptospermums can be grafted to *Kunzea flavescens*. I even grafted Geraldton wax to *Kunzea flavescens*. Also the time of the year grafted is important for a good success rate.

Save our Flora PowerPoint Presentation

Ready to go!

30 slides approx 30 mins. talk

**If you are interested in obtaining
this presentation
please email me**

**I can send it in an email (4.3MB)
or as a CD**

Send me a C5 stamped addressed envelope

Attach 2 stamps

or on a memory stick

**Send me a blank memory stick plus a stamped
addressed envelope - 2 stamps**

Permission to reproduce the Grafting article was obtained from Merv Hodge. It is the first of a series of articles I hope to include in our Bulletins from old volumes of Australian Plants.

In the 1960s and 70s enthusiasts could not obtain the large variety of Australian Plants now on the market. They were forced to learn to propagate and out of that generation grew a number of highly skilled propagators many of whom went on to start their own nurseries. They experimented and influenced the new generation of modern propagators and I feel that the value of their original work should not be lost but shared again.

I'm sure there might be articles in other journals which could also be shared. Please send me anything which you would like me to include.



**Office of
Environment & Heritage**
NSW National Parks & Wildlife Service



Our reference: MD14/5437

Ms Maria Hitchcock
maria.hitchcock@gmail.com

Dear Ms Hitchcock

I refer to your email to the Minister for the Environment, Rob Stokes MP, seeking advice on the application of Subdivision C - Listed threatened species and communities of the *Environment Protection and Biodiversity Conservation Act 1999* (EPBC Act). Your email was referred to the Office of Environment and Heritage (OEH) and I have been asked to reply.

Thank you for bringing your online initiative *Save our Flora* to my attention. It is always encouraging to hear about the efforts that passionate people in the community are making to help conserve our precious native flora and fauna. I hope the information below helps answer your queries.

No license is required to possess threatened species on your land

Land owners can 'possess' threatened species that are naturally occurring on the property, but cannot pick, hold for cultivation or sell them without a licence. There are separate licenses required for picking and cultivating threatened plant species and for selling them.

All sales of threatened plant species must be licensed or prove legal sourcing

It is legal to sell a threatened plant species where the stock plant originated in a garden only if it is sold under a Growers License issued under section 132 of the *National Parks and Wildlife Act 1974* (NPW Act).

Retailers do not need their own licence, unless they are also the producers of the material. A retailer must be able to prove legal sourcing and compliance with tag requirements. More information about plant licensing for commercial trade is available from the OEH website at <http://www.environment.nsw.gov.au/wildlifelicences/CommercialUseOfNativePlants.htm>.

Threatened species that have been legally sourced can be cultivated and traded without a license

It is not illegal to exchange the seed or cuttings of threatened plant species as long as the material is legally sourced. Legally sourced cultivated material can be propagated and traded to other parties to hold without a licence, provided there is no financial transaction.

Proving material is legally sourced requires evidence

The only defence to selling a threatened species is via a Growers licence issued under section 132 of the NPW Act (unless you are a retailer – see above). The legal sourcing of a parent material must be demonstrated prior to the issue of the licence.

PO Box A290 Sydney South NSW 1232
59-61 Goulburn St Sydney NSW 2000
Tel: (02) 9995 5000 Fax: (02) 9995 5999
TTY (02) 9211 4723
ABN 30 841 387 271
www.environment.nsw.gov.au

Contact us: saveourflora@gmail.com

Maria 02 6775 1139

Bob 02 6495 0306



Page 2

To prove that threatened plants or material purchased is legally sourced, evidence is required in the form of receipts for the purchase/acquisition of the material showing sufficient detail to trace the name and address of the seller. Plants must be appropriately tagged.

In terms of trading and swapping material that may have historic origins that predate the listing of the species under the *Threatened Species Conservation Act* (TSC Act), the proof required would likely take the form of horticulture records, if available, or a statutory declaration from the person(s) involved in obtaining and using that material.

In your example of *Boronia clavata*, a species listed under the EPBC Act, a licence is not required to possess or pick from propagated sources. As it is not native to NSW, wild sourcing is not applicable. A Growers licence is required to sell the species.

For a species that naturally occurs in NSW and is listed under the TSC Act, for example *Grevillea guthrieana*, a licence is required to harvest material from the wild for propagation purposes (via a threatened species or scientific licence), and to sell any propagated material (Growers licence). A licence is not required to possess the species if it occurs naturally on your property, or if you have sourced the species from appropriately licensed stock.

Whilst not directly related to your enquiry regarding listed threatened species under the EPBC Act, *The Commercial Harvest, Salvage and Propagation of Protected Whole Plants Sustainable Management Plan 2013-2017* (Management Plan) issued by OEHS further illustrates the management of commercial harvesting and the production of protected whole plants in NSW. You can find this Management Plan on the OEHS website at <http://www.environment.nsw.gov.au/wildlifelicences/wholeplantsmp.htm>.

If you have any questions regarding this issue, Ms Julie McInnes, Scientific Licensing Officer, NPWS, can be contacted by phone on 9585 6539 or by email at julie.mcinnnes@environment.nsw.gov.au.

Yours sincerely

MICHAEL WRIGHT
Acting Deputy Chief Executive
National Parks and Wildlife Service

Save our Flora

Seed and Cuttings Exchange

Please send all requests directly to the person making the offer.

Please follow the correct protocols for requests of seed or cuttings. These are detailed on the next page. Please note that some species are in very short supply and cutting material may be limited. Please note that in order to streamline this activity addresses will be published with the offers so that people can apply to the grower directly. Where there is no address please send your request to saveourflora@gmail.com

Maria Hitchcock

16 Hitchcock Lane Armidale NSW 2350
Correa eburnea
Callistemon pungens
Grevillea wilkinsonii
Zieria adenodonta

Arthur Baker

55 Moran ST Gatton Qld 4343
Gardenia psidiodes
Grevillea quadricauda
Grevillea glossadenia
Eucryphia wilkiei
Graptophyllum ilicifolium
Xanthostemon formosus
Phaius tancarvilleae
Plectranthus nitidus
Zieria prostrata
Grevillea mollis?
Eremophila nivea
Dodonaea rupicola
Xanthostemon arenaris
X verticulatus/seeds or cuttings
Kunzea flavescens
K granitica
Callistemon pearsonii
C flavovirens{seeds}
Melaleuca irbyana
Lilaeopsis brisbanica {Water plant}
Hernandia Bivalis
Spathoglottis Paulinia {Tropical ground orchid}
Rhododendron Lachiae

Charles Farrugia

Eremophila denticulata ssp trisulcata
Eremophila denticulata ssp denticulata
Eremophila nivea (blue form)
Eremophila nivea (white form) - limited.

Eremophila vernicosa – extremely limited – plant just recovering from a winter battering also I need to do some more grafts.

Russell Dahms

Boronia clavata

Denise & Graeme Krake

752 Warrigal Range Rd. Brogo NSW 2550
 Seed of
Hakea dohertyi
Hakea ochoptera
Hakea longiflora
Grevillea maccutcheonii, [this seed is still green]

Geoff & Gwynne Clarke

Grevillea humifusa - cuttings
Angophora robur - seed
Dodonaea crucifolia - cuttings or seed
 This was named a couple of years ago by Ian Telford who came down from Armidale to look over our block. Many people were calling it *Dodonaea hirsuta*, but it is not very hairy and has no hairs at all on the fruits. It also grows in a nearby flora reserve. If people would like to try this I can make it available when the material is ready. I have grown it successfully from cuttings, but it does not live long after planting out. It also produces seed and I can collect that after the next flowering (spring fruits). It grows happily around the block, popping up from seed here and there, produces plenty of seed, but it is not long lived even when self sown. Fruits are showy reds. I think it's worth a try.

Bob O'Neill

7 Hillsmeade Drive, Narre Warren South, Vic. 38051
 want to increase our range of Lechenaultias and *Correa pulchellas*. Can anyone help us out? Both of these groups of plants are doing well for us at Narre Warren South, Vic. I would be delighted to offer cuttings from our range to interested people. Some plants may be available to people who are able to come to our home address.

Paul Kennedy (Leader ANPSA Hakea SG)
 I am looking for seed or cuttings of *Hakea pedunculata* which grows naturally on Cape York near swamps. We have moved into our new home at 210 Aireys St. Elliminyt and have now begun the task of reintroducing all the Banksia and Hakea species.

Do you have any EPBC plants growing in your garden with sufficient foliage to share cuttings with our members? Let me know and I'll print it here. It

Save our Flora

Requesting and sending seed by post

Please follow these simple steps.

Make a request

1. Send your request by email first. It will be forwarded to the grower so you can request seed and ask for the address.
2. Send your request enclosing a self-addressed envelope with two 60c stamps attached. Post the envelope.

Send seed

1. When you receive an envelope with a seed request, package up the required seed which includes the name, provenance (if known) and date of collection. Add any tips on germinating the seed and post.

Receiving seed

1. Seed should be stored in paper (small manilla seed packets are best but any cheap envelopes will do) and kept in a cool dark place. Some people use those small paper lolly bags and staple them at the top. Add mothballs if you like. This will prevent insect attack. I save moisture absorbers from medicine bottles and add them to my seed drawer to ensure the seeds do not rot.

Seed life varies according to species. Acacias will last for many years while Flannel Flower needs to be really fresh. Old seed may not germinate and needs to be thrown out. Test some of your seed periodically. It's worth asking seed suppliers for the age of certain species of seed before purchasing.

Requesting and sending cuttings by post

Please follow these simple steps.

Make a request

1. Send your request by email first. It will be forwarded to the grower so you can request cuttings and ask for the address.
2. Purchase an Express Post small satchel for \$10.55. it will hold up to 500 gms.
3. Self address your satchel and place it in an envelope with your cuttings request. Add a label/s with the name of the species and sender. Pencil is best for writing on labels.
4. Post the envelope.

Send cuttings

1. When you receive an envelope with a satchel inside, cut about 6 stems of the requested species. The best time to do this is early morning. Store cuttings in the crisper part of the fridge until they are ready to be posted.
2. Wrap the cuttings in damp newspaper and place them in a cliplok plastic bag. Make sure you label each parcel with the names of the species and sender. Squeeze air out of the bag and fasten top.
3. Put the bag in the satchel and post.

Receiving cuttings

1. As soon as you receive your cuttings put the

Group Members

ANPSA Groups

APS Melton Bacchus Marsh Vic
 SGAP Ipswich Qld
 SGAP Sunshine Coast and Hinterland Qld
 APS Echuca Moama Vic

Botanic Gardens and Reserves

Hunter Regional Botanic Gardens
 Tamworth Regional Botanic Gardens
 Lindum Park Flora and Fauna Reserve
 Burrendong Arboretum Wellington

Nurseries

Bilby Blooms Binnaway NSW
 Cool Natives Nursery Armidale NSW
 Mole Station Native Nursery Tenterfield NSW

Seed Suppliers

Victorian Native Seeds

Study Groups

Acacia SG
 Correa SG
 Epacris SG
 Garden Design SG
 Grevillea SG
 Hakea SG
 Waratah & Flannel Flower SG

Do you belong to a group interested in growing or conserving native flora?

Why not ask them to join us?